MYCORRHIZAL ASSOCIATIONS IN EIGHT SELECTED TERRESTRIAL ORCHIDS FROM WESTERN GHATS OF GOA

M J Bukhari and D Velip

Department of Botany, Government College of Arts, Science and Commerce, Quepem- 403 705, Goa, India

Abstract

The present study was carried out to assess the mycorrhizal associations in the roots of eight terrestrial orchid species [Crepidium versicolor (Lindl.) Sushil K.Singh, Agrawala & Jalal, Eulophia picta (R.Br.) Ormerod, Habenaria crinifera Lindl., H. diphylla (Nimmo) Dalzell, H. marginata Colebr., Nervilia concolor (Blume) Schltr., Pecteilis gigantea (Sm.) Raf., and Peristylus plantagineus (Lindl.) Lindl.] from the Western Ghats of Goa. The present communication revealed the occurrence of mycorrhizal associations in the roots of eight selected terrestrial orchid species from Western Ghats of Goa. The colonization was characterized by the presence of pelotons in the root cortical cells.

Introduction

MYCORRHIZAL FUNGI form a mutualistic association with the roots of terrestrial orchids and are considered to be essential for orchid seed germination. These have a significant effect on the vegetative and reproductive growth of the orchid plants. Orchid mycorrhiza is the symbiotic association between fungi and epiphytic and terrestrial species of orchids (Dearnaley, 2007). The association is characterized by the entry of fungal hypahe in the parenchymatous cells of germinating seeds through trichomes and suspensor cells (Smith and Read, 2008) and also through root velamen and cortical cells at adult stage (Clements, 1988), resulting in the formation of intracellular coiled fungal hyphae called pelotons. These coiled fungal hyphal structures, the pelotons serve as a site of nutrient transfer between the symbionts (Dearnaley et al., 2016; Rasmussen, 1995; Selosse, 2014) essential for the perpetuation of orchids in their natural habitats. This symbiotic association has a significant effect on the vegetative and reproductive growth of plant (Pandey et al., 2013; Perotto et al., 2014).

The importance of this mutualistic association with mycorrhizal fungi for germination of orchid seeds and for successful seedling development in terrestrial orchids has been well documented in the earlier reports (Batty et al., 2001; Bhatti et al., 2017; Rasmussen, 2002). Literature studies revealed the occurrence of mycorrhizal colonization in many orchids (Peterson and Farquhar, 1994; Senthilkumar and Krishnamurthy, 1998; Vij and Sharma, 1983, 1988; Vij et al., 1995, 2002; Zettler, 1997). Studies have also revealed how orchids gain mineral nutrients and sometime even organic compounds from interact with orchid mycorrhizal fungi (Li et al., 2021). The distribution of orchids in Goa is

concentrated mainly in the Western Ghats region, which accounts for 86% of the total orchid diversity of the state (Jalal, 2019). The present communication revealed the occurrence of mycorrhizal associations in the roots of eight selected terrestrial orchids species from Western Ghats of Goa.

Material and Methods

Study Area

The present study was carried out at various localities from Goa *i.e.*, Surla, Gaoundongri, Bhupar-Gaoundongri, Dabhal, Dharbandora, Netravali, Cuncolim, Cotigao, and Pali.

Plant Identification

Plants collected during the present study were identified using flora of Goa (Rao, 1986) and Abraham and Vatsala (1981).

Sampling

During the present study, eight terrestrial orchid species belonging to six genera *i.e.*, *Crepidium*, *Eulophia*, *Habenaria*, *Nervilia*, *Pecteilis*, and *Peristylus* in flowering stage were collected from different localities in Goa (Table 1). Pseudobulb/rhizome of each of the species was carefully removed, stored in polyethylene bags and brought to the laboratory for further processing. The roots were freed from the adhering soil debris by washing under running tap water and preserved in 70% alcohol for microscopic observations.

Mycorrhizal Studies

For mycorrhizal studies, thin free hand sections of roots were taken and stained with 0.05% trypan blue (Phillips

Table. 1. Data regarding flowering period, locality and habitat of eight terrestrial orchids in Western Ghats of Goa.

Species	Flowering period	Locality	Habitat
Crepidium versicolor (Lindl.) Sushil K.Singh, Agrawala & Jalal	July-August	Bhupar-Gaoundongri	It is mainly found in evergreen forest floor under shade, moist places; it occurs at about 448 m elevation
Eulophia picta (R.Br.) Ormerod	June-September	Netravali	It is found in moist deciduous forest, grows in well-drained soil; it occurs at 102 m elevation
Habenaria crinifera Lindl.	August-September	Surla, Gaoundongri	Grows on well-drained soil with humidity of 70-80%; it occurs at 111 m elevation
H. diphylla (Nimmo) Dalzell	August-September	Cotigao	Grows on well-drained humus rich soil; it occurs at 98 m elevation
H. marginata Colebr.	September-November	Pali, Dabhal	The plant species found in open grasslands and the forest areas; it occurs at 72 m elevation
Nervilia concolor (Blume) Schltr.	May-June	Netravali	It is found in moist deciduous forest, grows in well-drained soil; it occurs at 102 m elevation
Pecteilis gigantea (Sm.) Raf.	September-October	Dabhal, Dharbandora, Cuncolim	It was found in open grassland, lateritic areas, sometimes found in evergreen forest patches; it occurs at 33 m elevation
Peristylus plantagineus (Lindl.) Lindl.	July-September	Gaoundongri, Netravali	It was found in between the gaps of semi-evergreen forest patches, with humus rich clay soils; it occurs at 130 m elevation

and Hayman, 1970) and were observed under microscope for studies on hyphae and peloton formation. Photographs of the transverse sections were taken using a digital camera of Samsung attached to the microscope.

Results and Discussion

All the selected eight terrestrial orchid species were found growing on varied habitats. Table 1 indicates the data regarding flowering period, locality and habitat of eight terrestrial orchids in Western Ghats of Goa. Crepidium versicolor was reported from higher elevation of evergreen forest floors. Eulophia picta was collected from moist deciduous forest of Netravali. Habenaria crinifera were found growing in well-drained humus-rich soil with humidity 70-80%, at 98 m elevation. H. diphylla was found growing on well-drained humus rich soil; it occurs at 98 m elevation. H. marginata was found growing in open grasslands. Nervilia concolor was collected from moist deciduous forest of Netravali. Pecteilis gigantea was found growing in open grassland, lateritic areas, sometimes found in evergreen forest patches as well while, Peristylus plantagineus was found between the gaps of semi-evergreen forest patches, with humus rich clay soils at 130 m elevation.

The results on the cross section of roots of all the eight species revealed the presence of pelotons and hyphae. Hyphal colonization was reported in the root hair, velamen tissues, and cortical region of the roots. In all the studied species, the hyphal entry was through the

root hair. Our results are in agreement with Senthilkumar and Krishnamurthy (1998) and Sathiyadash et al. (2012) where the fungal entry was through the root hair. Pelotons and digested pelotons were concentrated mainly in the cortical region (Fig. 1a-p). In the present study, the extent of hyphal coiling which is known to increase the inter-facial area between the symbionts varied amongst all the presently studied orchid species. Earlier studies reveal that peloton dimensions were related to cell dimension indicating that the size of pelotons seems to depend on the size of the host cell (Sathiyadash et al., 2012). In Crepidium versicolor and Eulophia picta, major part of cortical region in the cross section showed the presence of pelotons while in the other species, a smaller portion of cortical region was occupied by the pelotons. The pelotons appeared as loosely arranged fungal mycelial network of spherical and hexagonal balls in the parenchymatous cells of the cortical region. The present study revealed that the mycorrhizal fungus which resides in the cortical cells helps in absorbing minerals and other nutrients from the surrounding medium which is then utilized by the host by a process of digestion of the fungal cells within the host tissue. Similar observations were earlier made in other terrestrial species (Vij et al., 1995, 2002; Zettler, 1997). One of the important feature of the orchid mycorrhizae is the lysis of the pelotons (Peterson and Currah, 1990).

Mycorrhizal fungi are well known for the translocation of sugars and phosphates to the orchid plant (Richardson *et al.*, 1992). This type of fungal symbiotic

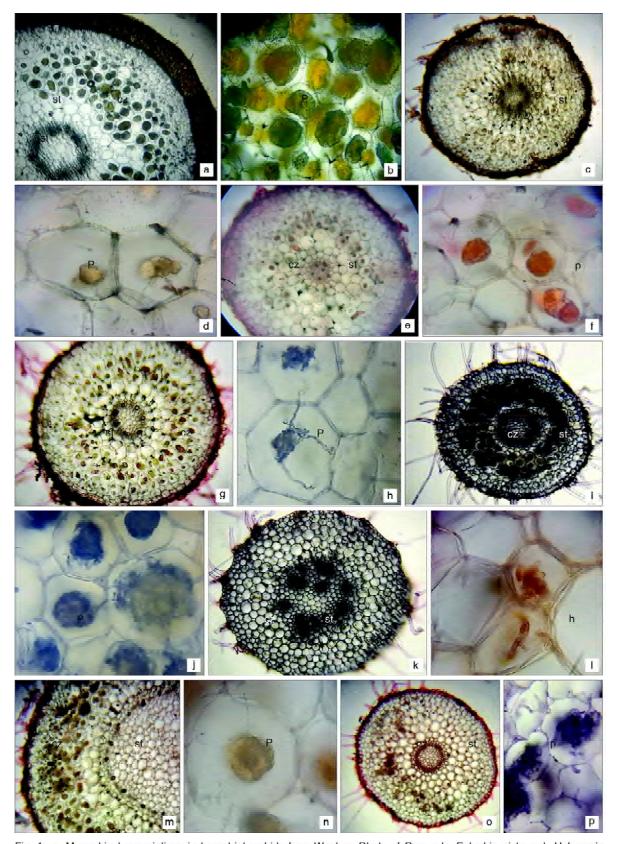


Fig. 1a-p. Mycorrhizal associations in terrestrial orchids from Western Ghats of Goa: a-b, *Eulophia picta*; c-d, *Habenaria crinifera*; e-f, *Habenaria diphylla*; g-h, *Habenaria marginata*; i-j, *Crepidium versicolor*, k-l, *Nervilia concolor*, m-n, *Pecteilis gigantea*; o-p, *Peristylus plantagineus* (cz, colonization zone; s-t, stele; p, pelotons; h, hyphae).

association has been reported earlier in terrestrial orchids (Jyothsna and Purushothama, 2013; Senthilkumar and Krishnamurthy, 1998; Vij and Sharma, 1983, 1988; Vij et al., 1995, 2002; Zettler, 1997). Studies by Yoder et al. (2000) has revealed the higher water content in the infected seedlings of both terrestrial and epiphytic orchids than non infected control and their implications in enhanced uptake. Germination is considerably enhanced with mycorrhizal fungi that facilitate the absorption of free water by their orchid seed host. Studies on successful germination of seeds with the aid of mycorrhizal fungi in terrestrial orchids has been studied earlier in some orchids (Bhatti et al., 2017; Rasmussen, 2002). Orchid mycorrhizal fungi have also been studied from other parts of the country. Earlier, Kaushik (1983) gave an illustrated account of intracellular mycorrhizal pelotons and fungal hyphae of 53 species of orchid roots with the help of transections and longitudinal sections. Kumar and Kaushik (2004) showed entrance of the fungus through root hair and observed pelotons in the cortical cells of roots of Zeuxine strateumatica and cultured and isolated the enzyme cellulase producing Rhizoctania solani. Kaushik and Pal (2011) described pathogenicity and symbiotic nature of the genus Rhizoctonia. Pal and Kaushik (2011) studied the antagonistic potential of Trichoderma viride to control Rhizoctonia solani both isolates from Rhynchostylis retusa and Aerides multiflora respectively. Subsequently Kaushik and Pal (2012) studied lignin degrading capacity of *Rhizoctania* solani isolated from Vanda testacea qualitatively and quantitatively.

During the present study, Rhizoctonia-like fungi were noted in the cross section of roots of all the studied species. Earlier studies have reported orchid association with a wide range of fungal species (Waud et al., 2017) and mostly with fungal taxa belonging to Rhizoctonialike species (Dearnaley et al., 2012; Jyothsna and Purushothama, 2013; Weiß et al., 2016). These mycorrhizal fungi play a significant role in rehabilitating threatened orchid species in their natural habitats (Dearnaley, 2007) as they are believed to ensure orchid survival in habitats which either are vulnerable to stressful conditions or these are with limited resources. Some of the selected species in the present study are of ethno-medicinal importance and call for conservation measures. The root paste of *Eulophia picta* is used for curing insect bites (Dash et al., 2008) and is an endangered orchid (Sheelavantmath et al., 2000). The fresh and dried rhizome of Crepidium versicolor is used by the tribal and local community of Goa for de-worming. Attempts have already been made towards conservation of some of the commercially important and rare. endangered, and threatened (RET) orchids by propagating them using tissue culture techniques (Anuprabha and Pathak, 2020; Anuprabha et al., 2017; Arora et al., 2016; Bhowmik and Rahman, 2023; Dhillon and Pathak, 2023; Hossain et al., 2012; Kaur et al., 2017; Kirti et al., 2023; Kumari and Pathak, 2021; Mutum et al., 2022; Pathak et al., 2022, 2023; Sunita et al., 2021; Thakur and Pathak, 2021; Tripura et al., 2022; Vasundhra et al., 2021).

To conclude, the present study provides evidence on occurrence of orchid mycorrhizal fungi in all the selected terrestrial species of Western Ghats of Goa. Knowledge of the mycorrhizal fungi is important for programmes aimed at the conservation and management of orchid species. Thus, isolation and identification of mycorrhizal fungi are important to understand the orchid-fungus relationship, which can lead to the development of efficient conservation strategies of the endangered and ethno-medicinally important terrestrial orchids occurring in Western Ghats of Goa.

References

- Abraham, A. and P. Vatsala. 1981. *Introduction to Orchids*. Tropical Botanic Garden and Research Institute, Trivandrum, Kerala, India.
- Anuprabha and Promila Pathak. 2020. Micropropagation of *Coelogyne fimbriata* Lindl. using pseudobulb explants. *J. Orchid Soc. India*, **34**: 131-36.
- Anuprabha, Promila Pathak, Ankush Prakash, and Jitender Kumar. 2017. Regeneration competence of *Dendrobium nobile* Lindl. through pseudobulb segments: A study *in vitro. J. Orchid Soc. India*, 31: 71-75.
- Arora, S. K., Promila Pathak, Shivani Verma, Ankush Prakash, Kriti Dhiman, and K. C. Mahant. 2016. Mass propagation of Dendrobium amoenum Wall. ex Lindl. through stem nodal explants: A study in vitro. J. Orchid Soc. India, 30: 51-55.
- Batty, A. L., K. W. Dixon, M. Brundrett, and K. Sivasithamparam. 2001. Constraints to symbiotic germination of terrestrial orchid seeds in a mediterranean bush-land. *New Phytol.*, **152**: 511-20.
- Bhatti, S. K., Jagdeep Verma, Jaspreet K. Sembi, and Promila Pathak. 2017. Symbiotic seed germination of *Aerides multiflora* Roxb.- A study *in vitro. J. Orchid Soc. India*, **31**: 85-91.
- Bhowmik, T. K. and M. M. Rahman. 2023. *In vitro* seed, seedling and SPSs development in *Habenaria digitata* Lindl. on different growth additives and PGRs supplemented MS medium. *J. Orchid Soc. India*, **37**: 59-67.
- Clements, M. A. 1988. Orchid mycorrhizal associations. *Lindleyana*, **3**: 73-86.
- Dash, P. K., S. Sahoo, and S. Ba. 2008. Ethnobotanical studies on orchids of Niyamgiri Hill Ranges, Orissa, India. *Ethnobot. Leafl.*, 12: 70-78.
- Dearnaley, J. D. W. 2007. Further advances in orchid mycorrhizal research. *Mycorrhiza*, **17**: 475-86.

- Dearnaley, J. D. W., F. Martos, and M. A. Selosse. 2012. Orchid mycorrhizas: Molecular ecology, physiology, evolution, and conservation Aspects. *In: Fungal Associations II: The Mycota IX* (ed. B. Hock) pp. 207-230. Springer-Verlag, Berlin, Heidelberg, Germany.
- Dearnaley, J. D. W., S. Perotto, and M. A. Selosse. 2016. Structure and development of orchid mycorrhizas. *In: Molecular Mycorrhizal Symbiosis* I (ed. F. Martin) pp. 63-86. John Wiley & Sons, Inc., Hoboken, U.S.A.
- Dhillon, M. K. and Promila Pathak. 2023. Asymbiotic seed germination in a medicinally important and near threatened terrestrial orchid, *Crepidium acuminatum* (D.Don) Szlach. from NorthWestern Himalayas: A study in vitro. J. Orchid Soc. India, 37: 49-57.
- Hossain, M. M., M. Sharma, and Promila Pathak. 2012. In vitro propagation of Dendrobium aphyllum (Orchidaceae)- Seed germination to flowering. J. Plant Biochem. Biotechnol., 22: 157-67.
- Jalal, J. S. 2019. Diversity and distribution of orchids of Goa, Western Ghats, India. J. Threat. Taxa, 11(15): 15015-42.
- Jyothsna, B. S. and K. B. Purushothama. 2013. Studies on the mycorrhiza of *Geodorum densiflorum* (Lam.) Schltr. from Western Ghats of Karnataka, India. *IOSR-JPBS.*, 6(5): 92-95.
- Kaur, S., Promila Pathak, Ankush Prakash, Anamika, and Aakanksha Sharma. 2017. Ex situ conservation of a floriculturally and medicinally important endangered orchid, Coelogyne cristata Lindl. J. Orchid Soc. India, 31: 15-22.
- Kaushik, P. 1983. Ecological and Anatomical Marvels of the Himalayan Orchids. Today and Tomorrow's Printers and Publishers, New Delhi, India.
- Kaushik, P. and P. Pal. 2011. *Rhizoctonia* a genus of orchid symbionts. *J. Orchid Soc. India*, **25**(1-2): 19-28.
- Kaushik, P. and P. Pal. 2012. Lignin degrading efficiency of Rhizoctonia solani: A study in vitro. J. Orchid Soc. India, 26(1-2): 79-82.
- Kirti, Promila Pathak, and K. C. Mahant. 2023. Asymbiotic seed germination and seedling development in commercially important and endemic orchids of Western Ghats, Aerides crispa Lindl.- A study in vitro. J. Orchid Soc. India, 37: 141-49.
- Kumar, R. and P. Kaushik. 2004. Isolation of a cellulase producing mycorrhizal fungus *Rhizoctonia solani* from *Zeuxine* strateumatica (Linn.) Schltr. J. Orchid Soc. India, 18(1-2): 101-08.
- Kumari, Anamika and Promila Pathak. 2021. De novo plantlet regeneration from leaf explants of Rhynchostylis retusa (L.) Blume: A study in vitro. J. Orchid Soc. India, 35: 47-53.
- Mutum, R. D., N. M. Chanu, T. N. Khanganba, and B. Thongam. 2022. Propagation and conservation of selected orchids of Manipur. J. Orchid Soc. India, 36: 95-101.
- Li, T., W. Yang, S. Wu, A. Selosse, and J. Gao. 2021. Progress and prospects of mycorrhizal fungal diversity in orchids. *Front. Plant Sci.*, **12**: 646325.
- Pal, P. and P. Kaushik. 2011. Antagonistic activity of *Trichoderma viride* against *Rhizoctonia solani* isolated from an orchid. *Vegetos*, **25**(2): 76-77.

- Pandey, M., J. Sharma, D. L. Taylor, and V. L. Yadon. 2013. A narrowly endemic photosynthetic orchid is non-specific in its mycorrhizal associations. *Mol. Ecol.*, 22: 2341-54.
- Perotto, S., M. Rodda, A. Benetti, F. Sillo, E. Ercole, and R. Balestrini. 2014. Gene expression in mycorrhizal orchid protocorms suggests a friendly plant fungus relationship. *Planta*, **239**: 1337-49.
- Pathak, Promila, Anamika Kumari, Brent D. Chandler, and Lawrence W. Zettler. 2023. In vitro propagation and phytochemical analysis of therapeutically endangered orchid, Vanda cristata Wall. ex Lindl. S. Afr. J. Bot., 153: 109-23.
- Pathak, Promila, Sunita, Anamika Kumari, Babita Thakur, Vasundhra, and Madhu. 2022. Regeneration competence of an endangered orchid, *Vanda cristata* Wall. ex Lindl. using leaf explants: A study *in vitro. S. Afr. J. Bot.*, **151**: 1018-24.
- Peterson, R. L. and R. S. Currrah. 1990. Synthesis of mycorrhizae between protocorms of *Goodera repens* (Orchidaceae) and Ceratobasidium cereale. Can. J. Bot., **68**: 1117-25.
- Peterson, R. L. and M. L. Farquhar. 1994. Mycorrhizas–integrated development between roots and fungi. *Mycologia*, **86**: 311-26.
- Phillips, J. M. and D. S. Hayman. 1970. Improved procedure for clearing root and staining parasitic and VA mycorrhizal fungi for rapid assessment of infection. *Trans. Br. Mycol. Soc.*, 55(1): 158-61.
- Rao, S. R. 1986. Flora of Goa, Daman and Diu, Dadra and Nagarhaveli. Vol II. Govt. of India, BSI Howrah, West Bengal, India.
- Rasmussen, H. N. 1995. Terrestrial Orchids: From Seed to Mycotrophic Plant. Cambridge University Press, Cambridge, U.K.
- Rasmussen, H. N. 2002. Recent developments in the study of orchid mycorrhiza. *Plant Soil*, 244: 149-63.
- Richardson, K. A., R. L. Peterson, and R. S. Currah. 1992. Seed reserves and early symbiotic protocorm development of *Platanthera hyperborea* (Orchidaceae). *Can. J. Bot.*, **70**: 291-300.
- Sathiyadasha, K., T. Muthukumar, E. Uma, and R. R. Pandey. 2012. Mycorrhizal association and morphology in orchids. *J. Plant Interact.*, **7**(3): 238-47.
- Selosse, M. A. 2014. The latest news from biological interactions in orchids: In love, head to toe. New Phytol., 202: 337-40.
- Senthilkumar, S. and K. V. Krishnamurthy. 1998 The role of root hair in the mycorrhizal association of the ground orchid *Spathoglottis plicata* Blume. *Mycorrhiza*, **10**(2): 15-17.
- Sheelavantmath, S. S., H. N. Murthy, A. N. Pyati, H. G. Ashok Kumar, and B. V. Ravishankar. 2000. *In vitro* propagation of the endangered orchid, *Geodorum densiflorum* (Lam.) Schltr. through rhizome section culture. *Plant Cell Tissue Organ Cult.*, 60: 151-54.
- Smith, S. E. and D. J. Read. 2008. *Mycorrhizal Symbiosis*. Academic Press, Cambridge, U.K.

- Sunita, Promila Pathak, and K. C. Mahant. 2021. Green pod culture of an endangered and medicinally important orchid, *Vanda cristata* Wall. ex Lindl. from Himachal Pradesh. *J. Orchid Soc. India*, **35**: 25-33.
- Thakur, Babita and Promila Pathak. 2021. Application of organic additives for the enhancement of seed germination and seedling development in an endangered and medicinal orchid, *Rhynchostylis retusa* (L.) Blume through asymbiotic culture. *J. Orchid Soc. India*, **35**: 99-107.
- Tripura, A., M. A. Sumi, T. K. Bhowmik, and M. M. Rahman. 2022. In vitro seed germination and phytochemical screening of an epiphytic medicinal orchid, *Pholidota imbricata* W. J. Hook. of Bangladesh. J. Orchid Soc. India, 36: 137-45.
- Vasundhra, Promila Pathak, and Anuprabha. 2021. *In vitro* asymbiotic seed germination and regeneration competence of leaf explants in *Satyrium nepalense* D.Don, a medicinally important, and an endangered terrestrial orchid of Kasauli Hills, Himachal Pradesh (NorthWestern Himalayas). *J. Orchid Soc. India*, **35**: 73-82.
- Vij, S. P. and M. Sharma. 1983. Mycorrhizal association in North Indian Orchidaceae: A morphological study. *Bibl. Mycol.*, 91: 467-503.
- Vij, S. P. and M. Sharma. 1988. Mycorrhizal endophytes of Nephelaphyllum BI. (Orchidaceae). In: International

- Conference on Research in Plant Science and its Relevance to Future. New Delhi, India.
- Vij, S. P., M. Sharma, and S. S. Datta. 1995. Mycorrhizal endophytes of *Spiranthes lancea* (Sw) Baker (Orchidaceae). *J. Indian Bot. Soc.*. 64: 175-79.
- Vij, S. P., T. N. Lakhanpal, and Ashish Gupta. 2002. Orchidoid mycorrhiza and techniques to investigate. *In: Techniques* in Mycorrhizal Studies (eds. K. G. Mukerji, C. Manoharachary, and B. P. Chamola) pp. 385-434. Kluwer Academic Publisher, The Netherlands.
- Waud, M., R. Brys, L. W. Van, B. Lievens, and H. Jacquemyn. 2017. Mycorrhizal specificity does not limit the distribution of an endangered orchid species. *Mol. Ecol.*, 26: 1687-701.
- Weiß, M., F. Waller, A. Zuccaro, and M. A. Selosse. 2016. Sebacinales-one thousand and one interactions with land plants. *New Phytol.*, **211**: 20-40.
- Yoder, J. A., L. W. Zettler, and S. L. Stewart. 2000. Water requirements of terrestrial and epiphytic orchid seeds and seedlings, and evidence for water uptake by means of mycotrophy. *Plant Sci.*, **156**: 145-50.
- Zettler, L. W. 1997. Orchid-fungal symbiosis and its value in conservation. *McIlvainea*, **13**: 40-45.