CONSERVATION, MANAGEMENT, AND UTILIZATION OF ORCHID GENETIC RESOURCES

L C De and Promila Pathak¹

ICAR-NRC for Orchids, Pakyong- 737 106, Sikkim, India

¹Department of Botany, Panjab University, Chandigarh- 160 014, UT, India

Abstract

Orchidaceae is one of the largest families of flowering plants and orchids are one of the most threatened plants due to habitat destruction and climate change, but many of them are also threatened by illegal harvest for horticulture, cultural use, food and medicine. In this review paper, trade related activities, economic importance of valuable orchid species and conservation measures are discussed in detail with a view to developing approaches that may allow us to address the threats on a broader scale to complement focused approaches for the species that are identified as being at the highest risk.

Introduction

ORCHIDACEAE IS one of the largest families of flowering plants (Chase et al., 2015; Willis, 2017) and are almost globally distributed. Till date, 28,484 species have been identified and accepted (Govaerts et al., 2017). By the end of 2017, the IUCN Global Red List included assessments for 948 orchid species, of which 56.5% are reported to be, threatened (IUCN, 2017). In addition to their geographical and taxonomic diversity, orchids are also widely used for a variety of reasons, both legally and illegally, sustainably and unsustainably (Fay, 2015a). One of the best-known plant groups in the global horticultural and cut flower trades (De, 2015; FloraHolland, 2015), orchids are also harvested, grown and traded for a variety of purposes, including as ornamental plants, medicinal products and food. Most popular global orchid trade is in artificially propagated cut flowers and plants grown under controlled conditions. During the period of 1996 to 2015, Taiwan and Thailand were the largest exporters, with most plants sent to South Korea (40%), the U.S.A (27%) and Japan (20%) (UNEP-WCMC, 2017).

Orchids are mostly vulnerable due to over-harvesting because many of them occur at low densities due to a variety of interacting factors such as recent speciation, specialized pollination mechanisms, habitat specificity, and the restricted distribution of mycorrhizal symbionts (McCormick and Jacquemyn, 2014; Swarts and Dixon, 2009a). The limited ecological studies on the conservation impacts of wild collection of epiphytic orchids indicate a low tolerance to harvest (Hu *et al.*, 2017; Mondragón, 2009).

Conservation Status

The family Orchidaceae is divided into five subfamilies (Apostasioideae, Cypripedioideae, Vanilloideae,

Received: August 19, 2018; Accepted: October 30, 2018

Orchidoideae, and Epidendroideae). Orchids account for nearly 8% of angiosperm species diversity (Chase et al., 2015; Willis, 2017). Major threats include habitat destruction, illegal harvesting, and because of their complex life histories, orchids are thought to be particularly vulnerable to the effects of global climate change (Fay and Chase, 2009; Gale et al., 2018; Swarts and Dixon, 2009a). All species of subfamily Cypripedioideae (the slipper orchids) were assessed for the Global Red List in a recent project, and due to a combination of habitat degradation and, in some cases, ruthless harvesting, nearly 90% of species were assessed and reported to be threatened (Fay and Rankou, 2016). Slipper orchids had expected a high level of threat, but even the family-wide figure of 56.5% showed that many orchid species are threatened of extinction. Their conservation should be regarded as urgent, if these unique plants are not to decline further.

Threats

Globally, both habitats and the species are under increasing pressure. There is also an increasing evidence that global climatic change may also be influencing species distribution (Fay, 2015b), and benefits and problems associated with assisted migration and translocations to climatically suitable localities are increasingly being discussed in relation to orchids (Ramsay and Dixon, 2003; Swarts and Dixon, 2009a). Like all plants, orchid species with their often complex interactions with pollinators, mycorrhizal fungi and host trees, are likely to be at a greater risk as they are dependent on other organisms that are also being affected by habitat or climatic change. Therefore, orchids face greater challenges than many other plant groups. Another aspect of threat relating to orchids is illegal harvesting; indiscriminate collection for horticultural purposes, has also been documented as having a major impact on some orchids, notably species of Cattleya, Laelia, Renanthera and some slipper orchids (Cypripedium, Paphiopedilum, Phragmipedium), and in some cases, these have been systematically stripped from the wild to the habitat of extinction. However, many orchids are not collected for horticulture or are collected in such small numbers that there is unlikely to be a much impact (Cribb et al., 2003; Fay, 2015a). As a result, these desirable orchids and because of perceived problems with their identification, all orchids were placed on the appendices of the Convention on International Trade in Endangered Species of Wild Fauna and Flora (CITES) (Cribb et al., 2003), and orchids account for more than 70% of the species listed on CITES. However, it is being observed that many orchid species are still being collected and transported across international borders, for use as medicine and food in addition to the horticultural trade, without the permits required under CITES (Fay, 2015a; Hinsley et al., 2018). The extent of the illegal trade is difficult to manage, but attempts are being made to estimate the extent of non-compliance with CITES regulations (Ghorbani et al., 2014; Hinsley et al., 2018). Notable examples of poorly documented trade relates to orchids collected for traditional medicine in East Asia and for production of the foodstuff, salep in the Eastern Mediterranean and the Middle East (de Boer et al., 2017; Kreziou et al., 2016) and Chikanda in South-Eastern Africa (Veldman et al., 2014); the development of novel DNA-based barcoding techniques (Huda et al., 2017, Ramudu and Khasim, 2016) is now opening up the opportunity to identify the orchid species in these processed foodstuffs. In addition, due to illegal trade, an unintended consequence of the listing of all species of orchid in CITES has led to the reduction in the collection of orchids for scientific purposes, including conservation research (Roberts and Solow, 2008).

Conservation Measures

About 70% of the world's orchids are epiphytic and/or lithophytic; 25% are terrestrial and 5% of the world's orchids grow in mixed substrates (lithophytic, epiphytic and terrestrial) (Arditti, 1992). These also occur as saprophytes. Pedersen *et al.* (2018) stressed the close link between collection based research and conservation. Approaches addressing conservation of process (rather than individual species) may be appropriate in groups which are relatively undergoing evolution due to hybridization and/or polyploidization (Ennos *et al.*, 2012). There are three prominent methods of conservation of genetic resources of orchid species namely, legislative measures, *in situ* conservation in Sanctuaries/Reserves, and *ex situ* conservation in Orchidaria/Botanic gardens by cultivation (Hegde, 2012).

Legislative Measures

Renanthera imschootiana (Red Vanda) and Vanda coerulea (Blue Vanda) are now included in schedule VI of Wildlife Protection Act (1972) of Government of India as amended in 1992. As a result, all orchids are protected plants under Wildlife Protection Act. The International Union for Conservation of Nature (IUCN) has a Species Survival Commission (SSC) with a well defined preservation programme for the Convention on International Trade in Endangered Species of Wild Flora and Fauna (CITES). Under this provision, orchids are treated as protected species. In India, three genera and eleven species are being treated as protected under Schedule-VI of Wild Life Protection Act, 1972 and simultaneously under CITES Appendix-I. All other species of India have been included in Appendix-II of CITES. The following species have been kept under CITES Appendix-I and Schedule-VI of Wild Life Protection Act of Govt. of India: Paphiopedilum charlesworthii, P. druryi, P. fairrieanum, P. hirsutissimum, P. insigne, P. spicerianum, P. venustum, P. wardii, Renanthera imschootiana, and Vanda coerulea. However, except P. druryi, which is reported from Kerala, all other species of Paphiopedilum belong to North East India. As per laws, no wild orchids can be traded with and so the plants listed above cannot be allowed for export. However, under CITES rules and regulations, whenever CITES Appendix-I species are cultivated, then these can be allowed for export, subject to condition that proper permit for possessing and growing these scheduled plants is obtained from the concerned State, provided the nursery is registered under Wild Life Preservation Office, Government of India.

Appendix-I includes threatened and extinct species. In fact, no trade in wild plants is allowed. Trade is allowed in cultivated and artificially propagated plants subject to licensing. Appendix-II covers species which may be threatened unless trade is strictly regulated. Infact, the whole family Orchidaceae is listed on Appendix-II. Trade in wild and propagated specimen is allowed subject to licensing. Further, all cultured orchid seedlings/plantlets in flasks or those aseptically raised from seeds and tissues are now exempted from CITES control w.e.f. April 16, 1993. These legislations have helped in checking illegal collection of orchids from the wild as non importing countries also accept plants of wild origin. In this regard, the Proceedings of the Seminar on CITES Implementation for Plants (1997), illustrates the method of recognizing the wild plants at the port and the procedure to deal with the same.

In Situ Conservation

It refers to the maintenance of the germplasm in its natural habitat allowing continual adaptation to the environment without any human interference. 2018)

Biosphere Reserves

These are versatile protected areas to preserve the genetic diversity in the representative ecosystem which are internationally recognized. The proposal for development of biosphere reserve was initiated by UNESCO in 1971 under the 'Man & Biosphere' (MAB) programme. The first biosphere reserve of the world was established in 1979. Presently, 564 biosphere reserves have been developed in 109 countries across the world. India has 17 biosphere reserves namely, Achanakamar-Amarkantak, Agasthymalai, Cold Desert, Dihang-Dibang, Dibru Saikhowa, Great Nicobar, Gulf of Mannar, Kachchh, Khangchendzonga, Manas, Nanda Devi, Nilgiri, Nokrek, Pachmarhi, Seshachalam Hills, Simlipal and Sunderbans for conservation of endemic, endangered and vulnerable orchid species.

National Parks

This is an area of adequate natural biological and geomorphological interest owned by a sovereign state having one or several ecosystems where conservation of wild life (both flora and fauna) is practiced along with educative and recreative interest, designated, created and protected by legislation. Hailey National Park, presently known as Jim Corbett National Park is the first developed National Park in India, in 1936. Presently, there are 98 National Parks in India. 96 different species of orchids are found in Simlipal National Park of Orissa and 150 different species of orchids are conserved in Buxa Tiger Reserve of West Bengal.

Sacred Groves

A sacred groove is a special type of area where all forms of life particularly the sacred tree species related to any particular culture are protected by a particular human community, race or tribe in the name of their respective deity. Himachal Pradesh, Karnataka, Kerala, Maharashtra, Andhra Pradesh, West Bengal and Chhattisgarh are very prominent states for sacred grooves. About 13,270 sacred grooves presently exist in India (Kumar *et al.*, 2016) and these may be important areas for *in situ* conservation of orchids of that particular locality.

Gene Sanctuary

Gene sanctuary is a protected area where broad spectrum of genetic variability is conserved to act as a reserve for future use and crop improvement. At present, India has 480 wildlife gene sanctuaries. Sessa Orchid Sanctuary of Arunachal Pradesh with 100 sq. km area conserves about 200 species of orchids. Similar types of sanctuaries have also been created in Sikkim at Deorali and Singtam.

Individual Trees

The epiphytic orchid species are conserved on tree species in their natural habitat where they attach themselves to the bark of trees, or the surface of other plants. Some of the host trees of orchid species are Lyonia ovalifolia, Benthamidia capitata, Quercus leucotricha, Diploknema butyracea, Berberis asiatica, Myrica esculenta, Castanopsis indica, Persea odoratissima, Pinus roxburghii, Rhododendron arboreum, Berberis cristata, Engelhardia spicata, Pyrus pashia, Shorea robusta, Mioromeles rhamnoides, Alnus nepalensis and Prunus cerasus (De et al., 2013). Their white thick roots are specially adapted to absorb moisture and dissolved nutrients. Because these tropical orchids usually grow high on the trees, rather than on the forest floor, they are accustomed to good air circulation and plenty of light.

Ex Situ Conservation

It refers to the preservation of germplasm outside the natural habitat. In India, Botanical Survey of India (BSI) is maintaining three National Orchidaria and Experimental Gardens, one each at Yercaud (Tamil Nadu), Howrah (West Bengal), and Shillong (Meghalya) where representative species of the region are being cultivated. Similarly, Arunachal Pradesh State Forest Research Institute is maintaining a large number of orchid species at Orchid Research Centre, Tipi, Itanagar, Sessa, Dirrang, Jenging and Roing as a measure of *ex situ* conservation of orchids. In Karnataka, three *ex situ* conservation centres have been established, one in Kodagu, another in Kudremukh and the third in Dhandeli (Rao and Sridhar, 2007).

Field Gene Banks

In this area, germplasm is collected from natural habitat or from other sources including commercial houses and nurseries and are maintained in the field or protected structures. In this connection, it is worthwhile to mention that in the field gene banks of TBGRI, Trivandrum, nearly 600 different species and 150 hybrids of orchids are maintained; NRC for Orchids, Pakyong, Sikkim has nearly 90 different genera and a number of hybrids of commercial orchids, and Orchid house at Panjab University, Chandigarh maintains nearly 100 species of orchids and some hybrids of commercial importance.

Botanical Gardens

These are protected areas where living plant specimens are conserved in fields or in protected structures providing significant information regarding mode of perpetuation, reproductive biology, taxonomical characters and propagation technique. At present, there are 13 botanical gardens in India maintaining a number of orchid species. About 43 species of orchids are collected and displayed in the orchid house of Lloyd Botanical garden, Darjeeling, West Bengal. Swarts and Dixon (2009b) focused on the role of botanic gardens in supporting orchid conservation scientifically and horticulturally.

Herbal Gardens

In these areas, medicinal plant genetic resources are reared in a protected area for maintaining them, generation after generation. Government of India has sanctioned funds for development of herbal gardens in 16 SAU's and research institutions in different agroclimatic regions of the country with a view to conserving and maintaining regional medicinal plants and endangered species (Gupta, 1993). In India, a networking among the herbal gardens is already developed at the Directorate of Medicinal and Aromatic Plants Research (DMAPR), where 83 such gardens with details of their species are registered.

Orchid Seed Gene Bank

Million of seeds are produced in a single capsule of orchid. However, they lack the functional endosperm and require specific mycorrhizal association for germination under natural conditions and consequently, the percentage of germination is low. Many orchids have been germinated through asymbiotic technique where germination is found as high as 90%. The seeds of orchids are orthodox in nature and provide a great scope for long term storage through low temperature.

In Vitro Conservation

This technique can be used for revitalization of orchid germplasm affected by virus and virus-like diseases through apical meristem culture, as a matter of fact, orchids are first plants to be tissue cultured, in this connection (Pritchard, 1989). Further, though attempts have been made to propagate orchid species in vitro using various explants (seeds, stem, root etc.) (Anuprabha et al., 2017; Arora et al., 2014, 2016; Bhattacharjee and Hossain, 2015; Bhatti et al., 2017; Borah et al., 2015; Chauhan et al., 2015; Hoque et al., 2016; Kaur and Pathak, 2014; Kaur et al., 2017; Pathak et al., 2016; 2017; Sibin and Gangaprasad, 2016; Sibin et al., 2014) so as to develop effective protocols for their *in vitro* propagation, the data is meager in terms size of the orchid family. There is a need for studies on genetic stability to avoid the somaclonal variants and slow growth cultures for longer storage duration to avoid frequent transfers.

Cryopreservation

Cryopreservation means long term storage or conservation of plant parts and reproductive materials at

a very low temperature, in the laboratory condition either in liquid nitrogen (-196°C) or in vapour phase (-150°C). Tissues/explants of orchids can be cryopreserved in liquid nitrogen cylinders as a long term storage procedure after proper treatment of cryoprotectants and plant vitrification solutions. In our country, National Bureau of Plant Genetic Resources (NBPGR) has created the facility of cryobank where 2.5 lakhs of germplasm lines can be stored (Singh, 2005).

Systematics and Conservation Genetics

Many groups of orchids are distributed in tropical regions, and phylogenetic studies are required so as to identify the number of species especially those that are phylogenetically isolated and consequently of high conservation value. Li et al. (2018) studied the use of phylogenetic measures as a mean for prioritizing members of Orchidaceae for conservation in the Indo-Burma Biodiversity Hotspot, revealing Thailand, South China and Vietnam as the areas with highest phylogenetic diversity and Tropidia curculigoides, Thaia saprophytica and Risleya atropurpurea as accounting for disproportionately great evolutionary distinctiveness. At the population level, genetic studies may be useful to identify regions or populations that should be treated as high priority for conservation. Earlier, development of markers was time consuming and expensive, but new technologies are speeding up marker development and allowing more loci to be studied than previously possible, and as a result, the quality of the information to be used in conservation planning will improve (Gargiulo et al., 2018).

Molecular genetic tool aids are used for species-level orchid identification. Techniques include Sanger sequencing-based DNA 'barcoding' techniques, which for plants typically compare two or more DNA regions (or 'markers') from each specimen with a library of verified reference samples (Hollingsworth et al., 2016). They also include the so-called next-generation sequencing techniques that use the whole genome or a much larger number of markers from across the entire genome to compare with a reference library. Barcoding approaches have been exploited for the monitoring of ornamental orchid trade (Phelps, 2015), so as to identify constituent species, in processed medicinal products (Wu et al., 2009; Yao et al., 2009), and most recently, to identify species in edible orchid products (Ghorbani et al., 2017; Veldman et al., 2017).

Conservation of Habitats

Habitat alteration, including total destruction, modification, and fragmentation, is widely recognised as the main threat to biodiversity in tropical regions where orchid diversity is the greatest. The relationship between epiphytic orchids and their host trees, calls for further research into the mechanisms controlling distribution of orchids on different species of trees (De et al., 2013; Rasmussen and Rasmussen, 2018). Some orchid conservation organisations such as the Orchid Conservation Alliance (OCA), which state that "preservation of natural orchid habitat preserves the orchids, their pollinators, their genetic diversity, and other fauna, as well as the birds, frogs, insects, reptiles, and mammals in the forests where they live" (OCA, 2017). A combination of creating new habitats, transplantation, and ex situ conservation in seed banks and living collections will meet these challenges. Conserving orchids in isolation from their pollinators, fungal associates and host plants means that the complexity of their biology is lost, even though the species still survives as has also been indicated earlier by Vanlalruati et al. (2016). For this reason, orchid conservationists suggest "integrated conservation", using ex situ techniques to support in situ conservation as an appropriate method.

Pollination Mechanisms, Pollinators, and Conservation

Orchids are popular for the wide range of pollination mechanisms and syndromes (Darwin, 1862; Micheneau et al., 2009) and the species diversity has been attributed, in part, to the diversity of pollen mechanisms (Cozzolino and Widmer, 2005). Because of the diversity of pollination mechanisms, Roberts (2003) revealed the importance of understanding pollination biology for effective orchid conservation, stating that "orchid conservation will require a case by case, functional ecosystem approach", and stated the need to conserve not only the orchid and the pollinator, but also in some cases the "pollinator food source, nesting site, larval host species, and in the case of parasitic pollinators, the larval host plant of its host species". Hutchings et al. (2018) studied that climate change can decouple the phenology of pollinator and orchid species, potentially leading to reproductive failure of the orchid. Recent papers have reviewed birds (Micheneau et al., 2006; van der Niet et al., 2015), crickets (Micheneau et al., 2010), fungus gnats (Phillips et al., 2014) and biting midges (Bogarín et al., 2018), as specialized pollinators. Many orchid species attract pollinators with the forms of deception including food deception, brood-site imitation, shelter imitation, rendezvous attraction and sexual deception (Buragohain and Chaturvedi, 2016; Jersáková et al., 2006), and recent discoveries of dual deceive (pseudopollen lacking food value; Davies et al., 2013), carrion mimicry (van der Niet et al., 2011), and production of fruit fly aggregation pheromones

(Karremans *et al.*, 2015) demonstrate that the understanding of the full complexities of orchid pollination is yet to be understood.

Mycorrhizal Associations and Conservation

Seed and protocorm development, including discussion of mycorrhizal associations and the survival of orchid seeds and plantlets in their natural habitats were reviewed by Yeung (2017). It is well known, the role of mycorrhizal fungi is crucial to the survival of selfsustaining populations of orchids (Hajong and Kapoor, 2016), but there is much research still to be conducted before understanding the mycorrhizal associations, especially with epiphytic orchid species. Even with temperate terrestrial species, the techniques including measurement of isotope enrichment of carbon, nitrogen and hydrogen now allow us to demonstrate the contribution that the fungi make to the nutrition of orchids, even when the plants are apparently capable of photosynthesis (Gebauer et al., 2016). Recently, techniques are used to demonstrate that reintroduced seedlings of Cypripedium calceolus had established mycorrhizal associations after planting out, despite being produced axenically (Fay et al., 2018).

Utilization of Genetic Resources

Horticultural Use

Over the last half a century, importance of orchids in floriculture has been amply highlighted by various workers (Hegde, 2014, 2016; Pathak et al., 2001) and agencies in India. These plants have been commercialized as ornamental plants in the horticultural and floricultural trade, which is unsurprisingly, dominated by species with attractive flowers, but it also includes species admired for their unusual growth habits (e.g. leafless orchids, such as species of Dendrophylax and Chiloschista), miniature size (e.g. species of Platystele and Bulbophyllum moniliforme), scent (e.g. species of Cattleya and Dendrochilum glumaceum), patterned leaves (e.g. jewel orchids in the genera Anoectochilus, Goodyera, Ludisia and Macodes) and as cut flowers (Renanthera imschootiana, Vanda coerulea). The principal orchid trade involves artificially propagated plants and cut flowers cultivated in commercial greenhouses. Reported Convention on the International Trade in Endangered Species of Wild Fauna and Flora (CITES) trade in live artificially propagated plants is dominated by a small number of genera with huge number of hybrids (e.g. Cymbidium, Dendrobium and Phalaenopsis, Cattleya, Oncidium). Orchids are consistently ranked among the best sellers in the global potted plant trade (FloraHolland, 2015; USDA, 2016) and

also has nearly 10% of all fresh cut flowers traded internationally (De, 2015). This represents an economically significant global trade, with exports of potted orchids from the Netherlands alone valued at almost 500 million in 2015 (FloraHolland, 2015). The largest areas of production are found in Thailand, Taiwan, The Netherlands and Japan, with demand for both potted and cut flowers growing in economic value annually (Griesbach, 2002; Hanks, 2015). Thailand, for example, sells roughly half of the orchids, it produces in the domestic market (Thammasiri, 2015). Important orchid genera used as potted plants in the international market are Ascocenda, Brassia, Cattleya, Cymbidium, Dendrobium, Epidendrum, Miltonia, Oncidium, Paphiopedilum, Phalaenopsis, and Vanda (Lopez and Runkle, 2005). Several local species of Ascocentrum, Calanthe, Cymbidium, Dendrobium, Paphiopedilum and Vanda etc. are in great demand in international market for breeding materials (Kumar and Sheela, 2007). They can also be preserved by drying for their use in flower arrangement and dried flower craft. These can be dried best using silica gel or borax for microwave drying, embedded drying or by freeze drying. Drying orchids is a challenging task as these flowers are considered difficult to be preserved. Dried orchids are used for different purposes such as these are used in vases and baskets and sometimes in shadow boxes. Bright flowers of orchid genera like Cattleya, Cymbidium, Dendrobium, Paphiopedilum, and Pholidota etc. can be used for drying (De et al., 2017).

Cultural and Ornamental Use

In Assam, the flowering spike of Rhynchostylis retusa, known as Kopou Phul is used by the girls to adorn their hair during the spring festival. The flowers of some other orchids like Coelogyne nitida and Vanda roxburghii are also used to adorn hair of girls of Assam and Arunachal Pradesh, in local festivals. The flowers of Papilionanthe teres are offered to Lord Buddha and spirits by the Khamtis and other Tai ethnics of Assam and Arunachal Pradesh. In Kameng district of Arunachal Pradesh, Dendrobium hookerianum, D. nobile and D. gibsonii are considered as the symbol of purity and sanctity by the local people. Monpas consider the flowers of Cymbidium grandiflorum important for holy worship. The young naga women of Manipur wear the orange flowers of Dendrobium densiflorum, behind their ears. Similarly, the flowers of Vanda coerulea are used by the women of Manipur, in hair during the autumn puja festival. In several countries, orchid species and hybrids are used as National Flowers. For example, Vanda Miss Joaquim in Singapore, Peristeria elata in Panama, and Lycaste skinneri var. alba in Guatemala. Orchids are depicted on stamps of several countries like Venezuela, USA,

New Zealand, Australia, Indonesia, India, Singapore, Japan, Russia, Thailand, Malaysia, and many others (Bhattacharjee and Das, 2008). As the orchids symbolize wealth, beauty and social status, orchid flower arrangements are used for good table decorations and venue decorations during weddings. Amongst orchids, Cymbidium, Dendrobium and Phalaenopsis are excellent for wedding counter pieces. An arch decorated with chic white silk combined with white orchids can be considered as an admirable orchid flower arrangement. In home, they can be displayed in three ways *i.e.*, single flower vases, plants in pots and traditional mixed flower arrangements. In Philippines and New Guinea, the stem of some Dendrobium species is used to make baskets and bracelets. In some tribes, Cattleya labiata var. autumnalis sap is used as glue for musical instruments. In Central America, the schomburgkias empty pseudobulbs are used to make horn (De and Pathak, 2015). Orchid flowers have historically been and continue to be traded for their ornamental value in a wide range of cultural and religious ceremonies. For example, flowers of Dendrobium maccarthiae are used as special temple offering in Sri Lanka, and flowers and pseudobulbs of species of Laelia are used in Mexican Day of the dead ceremonies (Duggal, 1971).

Use as Food

Orchids used for human consumption include globally important products, such as Vanilla flavourings (extracts of Vanilla), and other edible products used on national and regional levels. Leaves, tubers, and pseudobulbs of different species are used for edible purposes. Vanilla, a major spice crop and source of vanillin comes from Vanilla planifolia. Salep is made from the polysaccharide-rich tubers of wild orchids that were traded predominantly in Turkey as far back as 1850 (Landerer, 1850). After collection, the orchid tubers are boiled in water, milk or ayran (a yoghurt-based drink) to render the enzymes in them, inactive and prevent tubers from re-growing (Tamer et al., 2006). They are then dried and ground into a powder which is used to make the drink called salep and ice cream called maras dondurma (Kasparek and Grimm, 1999). At least 35 species of orchids are used to make salep, including species from the genera Anacamptis, Dactylorhiza, Himantoglossum, Ophrys, Orchis, Serapias and Steveniella (Kasparek and Grimm, 1999; Kreziou et al., 2016; Ghorbani et al., 2017). The tubers of terrestrial orchids are used in several african countries in the production of chikanda, a large cake with a meat-like structure, made of ground orchids and peanuts baked with ashes or baking soda (Bingham, 2009; Kaputo, 1996). Chikanda is a traditional dish eaten by the Bemba tribe in Northern Zambia (Richards, 1939) and by a tribe in the Katanga province 2018)

of the Democratic Republic of Congo (Malaisse and Parent, 1985), the Sumbawanga region in Tanzania (Davenport and Ndangalasi, 2003; Nyomora, 2005), Malawi (Kasulo et al., 2009) and the Bayam people in Cameroon, where the dish is prepared in a similar way with tubers of two species of Habenaria and is called napssié (Menzepoh, 2011). Predominant orchid species used for chikanda, generally belong to three genera *i.e.* Disa, Habenaria and Satyrium (Bingham and Smith, 2002; Bingham et al., 2003; Challe and Struik, 2008; Challe and Price, 2009; Davenport and Ndangalasi, 2003; Hamisy, 2007; Nyomora, 2005). Anoectochilus leaves are used as vegetables in Indonesia and Malaysia. Pseudobulbs of Cymbidium madidum and Dendrobium speciosum and tubers of Microtis uniflora and Caladenia carnea are eaten. The popular beverage called as Faham or Madagascar Tea on the islands of Mauritius and Madagascar is prepared from an orchid Jumellea fragrans. In Bhutan, the inflorescence or the flowers and pseudobulbs of Cymbidium spp. are eaten.

Medicinal Use

Many orchids are rich in alkaloids. Experimental evidences have reported the isolation of a number of alkaloids like anthocyanins, stilbenoids and triterpenoids from orchids. Orchinol, hircinol, cypripedin, jibantine, nidemin and loroglossin have been reported from orchids. Subsequently, orchids are also used in traditional medicine systems around the world, from subsistence to commercial levels. Some of the commercial medicinal uses of orchids include Chinese and South Asian Ayurvedic traditional medicine (Leon and Lin, 2017; Teoh, 2016). They are also known to be utilized in some African traditional medicine, e.g. Vanilla madagascariensis in Madagascar (Randriamiharisoa et al., 2015), North American folk medicine, e.g. Cypripedium acaule and C. parviflorum (Henkel, 1906) and the Unani medicine system, e.g. Dactylorhiza hatagirea, Vanda tessellata, Cymbidium bicolor and Ipsea speciosa (Khajuria et al., 2017; Thakur and Dixit, 2007). Medicinal orchids are also traded widely around the world, including Europe as various traditional medicines and health supplements (Brinkmann, 2014).

Use in Chinese Traditional Medicine

The medicinal value of orchids was reportedly first recognized in the 28th century BC by Shennong, China's founding emperor and patron deity of agriculture (Bulpitt, 2007; Hong, 2004). The most prominently cited orchids in Chinese Traditional Medicine are various *Dendrobium* spp. used to make the drug *shi-hu* from *D. catenatum*, *D. officinale*, *D. loddigesii*, *D. moniliforme* and *D. nobile* (Leon and Lin, 2017; Teoh, 2016). In addition, tubers of *Gastrodia elata* (from which *tian-ma* is prepared),

rhizomes of *Bletilla striata* (from which *bai-ji* is derived), the rhizomes and stems of *Anoectochilus* spp. (*jin-xianlian*) and the corms of *Cremastra appendiculata*, *Pleione bulbocodioides* and *P. yunnanensis* (from which *shan ci gu* is prepared) are all used (Leon and Lin, 2017; Teoh, 2016).

Use in Ayurvedic Medicine

The concept of Ayurvedic medicine originated in the Indian Subcontinent and has become globally practised, as part of complementary and alternative medicines. It includes a wide range of medicines, including Asthavarga preparations (Dhyani, et al., 2010) used to treat a variety of ailments. Nepal's Ayurvedic trade has been reported to involve nearly 94 orchid species (Acharya and Rokaya, 2010; Subedi et al., 2013), including Crepidium acuminatum, Habenaria intermedia, Herminium edgeworthii and Malaxis muscifera (Dhyani et al., 2010; Hossain, 2009; Khajuria et al., 2017). Eulophia spp. are also widely used medicinally across large parts of India [E. dabia, E. spectabilis (= E. nuda): Jalal et al., 2014], and D. hatagirea is used to treat a range of ailments (Pant and Rinchen, 2012). Estimates suggest that 6200-31,000 kg of D. hatagirea are harvested annually in the north-eastern Himalayan region of Sikkim (Rai et al., 2000; Unival et al., 2002), with each kilo having nearly100 individuals (Pant and Rinchen, 2012). Paphiopedilum druryi, an IUCN-listed Critically Endangered species endemic to Kerala, also continues to be collected for medicinal use and horticulture (Maridassa et al., 2008; Rankou and Kumar, 2015).

References

- Acharya, K. P. and M. B. Rokaya. 2010. Medicinal orchids of Nepal: Are they well protected? *Our Nature*, **8**: 82-91.
- Anuprabha, Promila Pathak, Ankush Prakash, and Jitender Kumar. 2017. Regeneration competence of *Dendrobium nobile* Lindl. through pseudobulb segments: A study *in vitro. J. Orchid Soc. India*, **31**: 71-75.
- Arditti, J. 1992. *Fundamentals of Orchid Biology*. John Wiley and Sons, New York, USA.
- Arora, S. K., Anuprabha, and Promila Pathak. 2014. Regeneration competence of *Arundina graminifolia* (D. Don.) Hochr. through stem disc culture: A study *in vitro. J. Orchid Soc. India*, 28: 109-13.
- Arora, S. K., Promila Pathak, Shivani Verma, Ankush Prakash, Kriti Dhiman, and K. C. Mahant. 2016. Mass propagation of *Dendrobium amoenum* Wall. ex Lindl. through stem nodal explants: A study *in vitro*. J. Orchid Soc. India, **30**: 51-55.
- Bhattacharjee, S. K. and S. P. Das. 2008. Orchids: Botany, Breeding, Cultivation, Uses and Post-Harvest Management. Aavishkar Publishers, Jaipur, Rajasthan, India.
- Bhattacharjee, D. K. and M. M. Hossain. 2015. Effect of plant growth regulators and explants on propagation on a monopodial

and sympodial orchid: A study *in vitro. J. Orchid Soc. India,* **29**: 91-102.

- Bhatti, S. K., Jagdeep Verma, Jaspreet K. Sembi, and Promila Pathak. 2017. Symbiotic seed germination of Aerides multiflora Roxb.- A study in vitro. J. Orchid Soc. India, 31: 85-91.
- Bingham, M. G. 2009. Chikanda, an unsustainable industry. *Pollinia,* **7**: 23-25.
- Bingham, M. G. and P. P. Smith. 2002. Zambia Plant Red List. In: Southern African Plant Red Data Lists (eds. J. S. Golding and S. O. Bandeira) pp. 140-56. Pretoria, South Africa.
- Bingham, M., G. M. Kokwe, and J. Golding. 2003. Chikanda: The edible orchids of Zambia. Veld Flora, 89: 15-17.
- Bogarín, D., M. Fernández, A. Borkent, A. Heemskerk, F. Pupulin, S. Ramírez, E. Smets, and B. Gravendeel. 2018. Pollination of *Trichosalpinx* (Orchidaceae: Pleurothallidinae) by biting midges (Diptera: Ceratopogonidae). *Bot. J. Linn. Soc.*, **186**: 510-43.
- Borah, N. J., S. Chakraborty, S. Roy Choudhary, and B. K. Dutta. 2015. *In vitro* propagation of *Paphiopedilum spicerianum* (Reichb. F.) Pfitz.- A rare and endangered orchid species from NorthEast India. *J. Orchid Soc. India*, **29**: 85-90.
- Brinkmann, J. A. 2014. Quick scan of Orchidaceae species in European commerce as components of cosmetic, food and medicinal products. PC22 Doc. 22.1 Annex. Available at: https://cites.org/eng/com/pc/22/index.php.
- Bulpitt, C. J. 2007. The use of orchids in Chinese medicine. *J. R. Soc. Med.*, **100**: 558-63.
- Buragohain, B. and S. K. Chaturvedi. 2016. Deceptive pollination in an endangered orchid, Vanda coerulea Griff. ex Lindl. (Orchidaceae). J. Orchid Soc. India, 30: 31-35.
- Challe, J. F. and L. L. Price. 2009. Endangered edible orchids and vulnerable gatherers in the context of HIV/AIDS in the southern highlands of Tanzania. J. Ethnobiol. Ethnomed., 5: 41.
- Challe, J. F. X. and P. C. Struik. 2008. The impact on orchid species abundance of gathering their edible tubers by HIV/AIDS orphans: A case of three villages in the Southern Highlands of Tanzania. NJAS-Wageningen J. Life Sci., 56: 261-79.
- Chauhan, Shaveta, Promila Pathak, Anuprabha, and Sanjay Sharma. 2015. Regeneration of *Eulophia dabia* through rhizome explants and flowering: A study *in vitro. J. Orchid Soc. India*, **29**: 61-65.
- Chase, M. W., K. M. Cameron, J. V. Freudenstein, A. M. Pridgeon, G. Salazar, C. Berg, and A. Schuiteman. 2015. An updated classification of Orchidaceae. *Bot. J. Linn. Soc.*, **177**: 151-74.
- CITES. 2017. Convention on International Trade in Endangered Species of Wild Fauna and Flora. Appendices I and II. Available: http://www.cites.org/eng/app/appendices.php.
- Cozzolino, S. and A. Widmer. 2005. Orchid diversity: An evolutionary consequence of deception. *Trends Ecol. Evol.*, **20**: 487-94.

- Cribb, P. J., S. P. Kell, K. W. Dixon, and R. L. Barrett. 2003. Orchid conservation: A global perspective. *In: Orchid Conservation* (eds. K. W. Dixon, S. P. Kell, R. L. Barrett, and P. J. Cribb). Kota Kinabalu: Natural History Publications, Borneo.
- Darwin, C. 1862. On the various contrivances by which British and foreign orchids are fertilised by insects, and on the good effect of intercrossing. John Murray, London.
- Davenport, T. R. B. and H. J. Ndangalasi. 2003. An escalating trade in orchid tubers across Tanzania's southern highlands: Assessment, dynamics and conservation implications. *Oryx*, **37**: 55-61.
- Davies, K. L., M. Stpiczyńska, and M. Kamińska. 2013. Dual deceit in pseudopollen-producing *Maxillaria s.s.* (Orchidaceae: Maxillariinae). *Bot. J. Linn. Soc.*, **173**: 744-63.
- de Boer, H. J., A. Ghorbani, V. Manzanilla, A. C. Raclariu, A. Kreziou, S. Ounjai, M. Osathanunkul, and B. Gravendeel. 2017. DNA metabarcoding of orchid derived products reveals widespread illegal orchid trade. *Proc. R. Soc. B, Biol. Sci.*, 284(1863): art. no. 20171182.
- De, L. C. 2015. Commercial Orchids. Berlin: De Gruyter Open. Available at: http://www.degruyter.com/view/product/ 456245.
- De, L. C. and Promila Pathak. 2015. Value Addition in Orchids. J. Orchid Soc. India, 29: 31-37.
- De, L. C., Geetamani Chhetri, and R. P. Medhi. 2013. Orchids- A wonderful crop for diversification. J. Orchid Soc. India, 27(1-2): 1-8.
- De, L. C., D. R. Singh, and Suman Thapa. 2017. Drying Technologies in orchids. *Inter. J. Environ. Sci. Nat. Res.*, 5(1): 1-5.
- Dhyani, A., B. P. Nautiyal, and M. C. Nautiyal. 2010. Importance of Ashtavarga plants in traditional systems of medicine in Garhwal, Indian Himalaya. *Inter. J. Biodivers. Sci. Ecosys. Ser. Manage.*, 6: 13-19.
- Duggal, S. C. 1971. Orchids in human affairs (a review). Q. J. Crude Drug Res., 11: 1727-34.
- Ennos, R. A., R. Whitlock, M. F. Fay, B. Jones, L. E. Neaves, R. Payne, I. Taylor, N. de Vere, and P. M. Hollingsworth. 2012. Process-based species action plans: An approach to conserve contemporary evolutionary processes that sustain diversity in taxonomically complex groups. *Bot. J. Linn. Soc.*, **168**: 194-203.
- Fay, M. F. 2015a. Undocumented trade in species of Orchidaceae. https://cites.org/sites /default/files /eng/com/pc/22/Inf/E-PC22-Inf-06.pdf.
- Fay, M. F. 2015b. British and Irish orchids in a changing world. *Curtis's Bot. Mag.*, **32**: 3-23.
- Fay, M. F. and M. W. Chase. 2009. Orchid biology- from Linnaeus via Darwin to the 21st century. *Ann. Bot.*, **104**: 359-64.
- Fay, M. and H. Rankou. 2016. Slipper orchids on the IUCN Red List. In: 2015 Annual Report to the Environment Agency-Abu Dhabi. Framework Support for Implementing the Strategic Plan of the IUCN Species Survival Commission. pp. 106-111. Abu Dhabi, UAE.

2018)

- Fay, M. F., M. Feustel, C. Newlands, and G. Gebauer. 2018. Inferring the mycorrhizal status of introduced plants of *Cypripedium calceolus* (Orchidaceae) in northern England using stable isotope analysis. *Bot. J. Linn. Soc.*, **186**: 587-90.
- FloraHolland. 2015. Facts & figures. *In: FloraHolland facts and figures 2015.* Available at: https://www.royalfloraholland. com/media/5685262/RoyalFloraHolland_Annual_Report_2015_ENG_facts_and_figures.pdf.
- Gale, S. W., G. A. Fischer, P. J. Cribb, and M. F. Fay. 2018. Orchid conservation: Bridging the gap between science and practice. *Bot. J. Linn. Soc.*, **186**: 425-34.
- Gargiulo, R., A. Ilves, T. Kaart, M. F. Fay, and T. Kull. 2018. High genetic diversity in a threatened clonal species, *Cypripedium calceolus* (Orchidaceae), enables long-term stability of the species in different biogeographical regions in Estonia. *Bot. J. Linn. Soc.*, **186**: 560-71.
- Gebauer, G., K. Preiss, and A. C. Gebauer. 2016. Partial mycoheterotrophy is more widespread among orchids than previously assumed. *New Phytol.*, **211**: 11-15.
- Ghorbani, A., B. Gravendeel, S. Zarre, and H. de Boer. 2014. Illegal wild collection and international trade of CITES-listed terrestrial orchid tubers in Iran. *Traffic Bull.*, 26: 52-58.
- Ghorbani, A., B. Gravendeel, S. Selliah, S. Zarre, H. de Boer. 2017. DNA barcoding of tuberous Orchidoideae: A resource for identification of orchids used in Salep. *Mol. Ecol. Res.*, **17**: 342-52.
- Govaerts, R., P. Bernet, K. Kratochvil, G. Gerlach, G. Carr, P. Alrich, A. M. Pridgeon, J. Pfahl, M. A. Campacci, D. Holland Baptista, H. Tigges, J. Shaw, P. Cribb, A. George, K. Kreuz, and J. J. Wood. 2017. *World Checklist of Orchidaceae*. Royal Botanic Gardens, Kew, London, UK.
- Griesbach, R. J. 2002. Development of *Phalaenopsis* orchids for the mass-market. *In: Trends in New Crops and New Uses* (eds. J. Janick and A. Whipkey) pp. 458-65. ASHS Press, Alexandria, Virginia.
- Gupta, R. 1993. Conservation and utilization of Indian medicinal plants. Indian J. Plant Gen. Res., 6: 131-38.
- Hajong, S. and R. Kapoor. 2016. Orchid mycorrhizal symbiosis: Evolution, molecular mechanism and role in orchid distribution. J. Orchid Soc. India, 30: 19-30.
- Hamisy, W. C. 2007. Orchid conservation project Tanzania. http://www.rufford.org/files/2-22.02.06%20 Detailed%20Final%20Report.pdf.
- Hanks, F. 2015. Orchid Cut Flower Industry. Report for the UK Agriculture and Horticulture Development Board and National Cut Flower Centre. https://horticulture.ahdb.org.uk/ sites/default/files/u3089/A%20review%20 of%20cutflower%20and%20foliage%20production%20statistics% 202015_0.pdf.
- Hegde, S. N. 2012. *Ex situ* and *in situ* conservation of orchids in India. *J. Orchid. Soc. India*, **26**(1-2): 1-4.
- Hegde, S. N. 2014. Status of exotic orchid hybrids and species in India: Its impact on Indian orchid industry. J. Orchid. Soc. India, 28: 23-29.

- Hegde, S. N. 2016. Commercial potential of orchids in India and societal benefits. *J. Orchid. Soc. India*, **30**: 43-49.
- Henkel, A. 1906. Wild Medicinal Plants of the United States (No. 89). US Department of Agriculture, Bureau of Plant Industry. Government Printing Office, Washington, D.C., USA.
- Hinsley, A., H. J. de Boer, M. F. Fay, S. W. Gale, L. M. Gardiner, R. S. Gunasekara, P. Kumar, S. Masters, D. Metusala, D. L. Roberts, S. Veldman, S. Wong, and J. Phelps. 2018. A review of the trade in orchids, and its implications for conservation. *Bot. J. Linn. Soc.*, **186**: 435-55.
- Hollingsworth, P. M., D. Z. Li, M. van der Bank, and A. D. Twyford. 2016. Telling plant species apart with DNA: From barcodes to genomes. *Philos. Trans. R. Soc. Lond. B: Biol. Sci.*, **371**: 20150338.
- Hong, F. F. 2004. History of medicine in China- when medicine took an alternative path. *McGill J. Med.*, **8**: 79-84.
- Hossain, M. M. 2009. Traditional therapeutic uses of some indigenous orchids of Bangladesh. *Med. Aromat. Plant Sci. Biotechnol.*, 42: 101-06.
- Hoque, M. M., L. Khaleda, and M. Al-Forkan. 2016. Development of cryopreservation protocols for five indigenous medicinal orchids of Bangladesh. J. Orchid Soc. India, 30: 105-10.
- Hu, A. Q., S. W. Gale, P. Kumar, R. M. K. Saunders, M. Sun, and G. A. Fischer. 2017. Preponderance of clonality triggers loss of sex in *Bulbophyllum bicolor*, an obligately outcrossing epiphytic orchid. *Mol. Ecol.*, 26: 3358-72.
- Huda, M. K., A. Price, and C. C. Wilcock. 2017. Identification of medicinal orchids of Bangladesh: DNA barcoding vs. traditional taxonomy. J. Orchid Soc. India, 31: 33-40.
- Hutchings, M. J., K. M. Robbirt, D. L. Roberts, and A. J. Davy. 2018. Vulnerability of a specialized pollination mechanism to climate change revealed by a 356-year analysis. *Bot. J. Linn. Soc.*, **186**: 498-509.
- IUCN. 2017. The IUCN Red List of Threatened Species, Version 2016-3. Available at: www.iucnredlist.org.
- Jalal, J. S., J. Jayanthi, and P. Kumar. 2014. Eulophia spectabilis: A high value medicinal orchid under immense threat due to overexploitation for medicinal use in Western Ghats, Maharastra. MIOS J., 15: 9-15.
- Jersáková, J., S. D. Johnson, and P. Kindlmann. 2006. Mechanisms and evolution of deceptive pollination in orchids. *Biol. Rev.*, 81: 219-35.
- Kaputo, M. T. 1996. The role of ashes and sodium bicarbonate in a simulated meat product from chikanda tuber (*Satyriasiva*). *Food Chem.*, **55**: 115-19.
- Karremans, A. P., F. Pupulin, D. Grimaldi, K. K. Beentjes, R. Butôt, G. E. Fazzi, K. Kaspers, J. Kruizinga, P. Roessingh, E. F. Smets, and B. Gravendeel. 2015. Pollination of *Specklinian* by nectar-feeding *Drosophila*: The first reported case of a deceptive syndrome employing aggregation pheromones in Orchidaceae. *Ann. Bot.*, **116**: 437-55.
- Kasparek, M. and U. Grimm. 1999. European trade in Turkish salep with special reference to Germany. *Econ. Bot.*, 53: 396-406.

- Kasulo, K., L. Mwabumba, and C. Munthali. 2009. A review of edible orchids in Malawi. J. Hortic. For., 1: 133-39.
- Kaur, S. and Promila Pathak. 2015. Reversion of reproductive phase to vegetative phase in the inflorescence segments of Saccolabium papillosum Lindl.: A study in vitro. J. Orchid Soc. India, 29: 75-79.
- Kaur, S., Promila Pathak, Ankush Prakash, Anamika, and Aakanksha Sharma. 2017. *Ex situ* conservation of floriculturally and medicinally important endangered orchid, *Coelogyne cristata* Lindl. *J. Orchid Soc. India*, **31**: 15-22.
- Khajuria, A. K., G. Kumar, and N. S. Bisht. 2017. Diversity with ethnomedicinal notes on orchids: A case study of Nagdev forest range, Pauri Garhwal, Uttarakhand, India. J. Med. Plants, 5: 171-74.
- Kreziou, A., H. De Boer, and B. Gravendeel. 2016. Harvesting of salep orchids in north-western Greece continues to threaten natural populations. *Oryx*, **50**: 393-96.
- Kumar, K. Madhu and V. L. Sheela. 2007. Status of breeding in orchids-A review. J. Ornament. Hort., 10: 199-208.
- Kumar, D., P. Kumari, S. S. Samant, and S. Paul. 2016. Assessment of orchid diversity in selected Sacred Groves of Kullu District, Himachal Pradesh, India. J. Orchid Soc. India, 30(1-2): 89-95.
- Landerer, X. 1850. Naturgeschichte und Pharmakognosie. Beiträge zur Pharmakognosie, Ueber Salep und die Salepisiden. *Archiv der Pharmazie*, **112**: 177-80.
- Leon, C. and L. Y. Lin. 2017. *Chinese Medicinal Plants, Herbal Drugs and Substitutes: An Identification Guide.* Kew Publishing, Kew, London, UK.
- Li, J., S. W. Gale, P. Kumar, J. Zhang, and G. Fischer. 2018. Prioritizing the orchids of a biodiversity hotspot for conservation based on phylogenetic history and extinction risk. *Bot. J. Linn. Soc.*, **186**: 473-97.
- Lopez, R. G. and E. S. Runkle. 2005. Environmental physiology of growth and flowering of orchids. *Hort. Sci.*, 40(7): 1969-73.
- Malaisse, F. and G. Parent. 1985. Edible wild vegetable products in the Zambezian woodland area: A nutritional and ecological approach. *Ecol. Food Nutri.*, **18**: 43-82.
- Maridassa, M., M. I. Zahir Hussain, and G. Raju. 2008. Phytochemical survey of orchids in the Tirunelveli Hills of south India. *Ethnobot. Leaflets*, **12**: 705-12.
- McCormick, M. K. and H. Jacquemyn. 2014. What constrains the distribution of orchid populations? *New Phytol.*, **202**: 392-400.
- Menzepoh, S. B. 2011. Les orchidées comestibles chez le people Bagam, au Cameroun. *Biotechnol. Agron. Soc. Environ.*, **15**: 509-14.
- Micheneau, C., J. Fournel, and T. Pailler. 2006. Bird pollination in an angraecoid orchid on Reunion Island (Mascarene Archipelago, Indian Ocean). *Ann. Bot.*, **97**: 965-74.
- Micheneau, C., S. D. Johnson, and M. F. Fay. 2009. Orchid pollination: From Darwin to the present day. *Bot. J. Linn. Soc.*, **161**: 1-19.

- Micheneau, C., J. Fournel, B. H. Warren, S. Hugel, A. Gauvin-Bialecki, T. Pailler, D. Strasberg, and M. W. Chase. 2010. Orthoptera, a new order of pollinator. *Ann. Bot.*, **105**: 355-64.
- Mondragón, D. 2009. Population viability analysis for *Guarianthe aurantiaca*, an ornamental epiphytic orchid harvested in southeast México. *Plant Species Biol.*, **24**: 35-41.
- Nyomora, A. M. S. 2005. Distribution and abundance of the edible orchids of the Southern Highlands of Tanzania. *Tanzania J. Sci.*, **31**: 45-54.
- O. C. A. 2017. The orchid conservation alliance. https:// orchidconservationallianc e.org/.
- Pant, S. and T. Rinchen. 2012. *Dactylorhiza hatagirea*: A high value medicinal orchid. *J. Med. Plants Res.*, **6**: 3522-24.
- Pathak, Promila, R. N. Sehgal, N. Shekhar, M. Sharma, and A. Sood. 2001. *Orchids: Science and Commerce*. Bishen Singh Mahendra Pal Singh, Dehradun, India.
- Pathak, Promila, Shivani Verma, Ankush Prakash, and K. C. Mahant.
 2017. Regeneration competence of an ornamentally important epiphytic orchid, *Rhynchostylis gigantea* (Lindl.)
 Ridl. through leaf segments: A study *in vitro. J. Orchid Soc. India*, **31**: 97-101.
- Pathak, Promila, Sanjeev K. Arora, Shivani Verma, Kriti Dhiman, K. C. Mahant, and Raja Jeet. 2016. Mass propagation of a floriculturally and medicinally important epiphytic orchid *Dendrobium amoenum* Wall. ex Lindl. through asymbiotic seed culture: A study *in vitro*. *Pb. Univ. Res. J. (Sci).*, 66: 39-45.
- Pedersen, H., G. Petersen, and O. Seberg. 2018. On the "Seidenfaden collection" and the multiple roles botanical gardens can play in orchid conservation. *Lankesteriana*, 18(1):1-12.
- Phelps, J. 2015. A Blooming Trade: Illegal Trade of Ornamental Orchids in Mainland Southeast Asia (Thailand, Lao PDR, Myanmar). TRAFFIC, Selangor, Malaysia.
- Phillips, R. D., D. Scaccabarozzi, B. A. Retter, C. Hayes, G. R. Brown, K. W. Dixon, and R. Peakall. 2014. Caught in the act: Pollination of sexually deceptive trap-flowers by fungus gnats in *Pterostylis* (Orchidaceae). *Ann. Bot.*, **113**: 629-41.
- Pritchard, H. W. 1989. Modern Methods in Orchid conservation: The Role of Physiology, Ecology and Management. Cambridge University Press, Cambridge, UK.
- Rai, L. K., P. Prasad, and E. Sharma. 2000. Conservation threats to some important medicinal plants of the Sikkim Himalaya. *Biol. Conserv.*, 93: 27-33.
- Ramudu, J. and S. M. Khasim. 2016. DNA barcoding of some Indian Coelogyne (Epidendroideae, Orchidaceae). J. Orchid Soc. India, 30: 65-73.
- Ramsay, M. M. and K. W. Dixon. 2003. Propagation science, recovery and translocation of terrestrial orchids. *In: Orchid Conservation* (eds. K. W. Dixon, S. P. Kell, R. L. Barrett, and P. J. Cribb) pp. 259-88. Sabah Natural History Publications, Kota Kinabalu, Malaysia.
- Randriamiharisoa, M. N., A. R. Kuhlman, V. Jeannoda, H. Rabarison, N. Rakotoarivelo, T. Randrianarivony, F. Raktoarivony, A.

Randrianasolo, and R. W. Bussmann. 2015. Medicinal plants sold in the markets of Antananarivo, Madagascar. *J. Ethnobiol. Ethnomed.*, **11**: 60.

- Rankou, H. and P. Kumar. 2015. Paphiopedilum druryi. The IUCN Red List of Threatened Species 2015:e.T15051138A15054919. http://dx.doi.org/10.2305/ IUCN.UK.2015-2. RLTS.T15051138A15054919.en.
- Rao, T. A. and S. Sridhar. 2007. *Wild Orchids of Karnataka- A Pictorial Compendium*, INCERT, Bangalore, India.
- Rasmussen, H. N. and F. N. Rasmussen. 2018. The epiphytic habitat on a living host: Reflections on the orchid-tree relationship. *Bot. J. Linn. Soc.*, **186**: 456-72.
- Richards, A. I. 1939. Land, Labour and Diet: An Economic Study of the Bemba Tribe. Oxford University Press, London, UK.
- Roberts, D. L. 2003. Pollination biology: The role of sexual reproduction in orchid conservation. *In: Orchid Conservation* (eds. K. W. Dixon, S. P. Kell, R. L. Barrett, and P. J. Cribb) pp. 113-36. Sabah Natural History Publications, Kota Kinabalu, Malaysia.
- Roberts, D. L. and A. R. Solow. 2008. The effect of the convention on international trade in endangered species on scientific collections. *Proc. R. Soc. Lond. B: Biol Sci.*, 275: 987-89.
- Sibin, N. T. and A. Gangaprasad. 2016. Development of *in vitro* propagation protocol for rapid and mass propagation of *Coelogyne nervosa* A. Rich., an endemic orchid of the Southern Western Ghats using immature seeds. *J. Orchid Soc. India*, **30**: 37-41.
- Sibin, N. T., A. Gangaprasad, and S. Anjusha. 2014. Effects of different organic additives on *in vitro* asymbiotic seed germination of *Arundina graminifolia* (D. Don) Hochr., an exquisite rare orchid. J. Orchid Soc. India, 28: 61-66.
- Singh, B. M. 2005. Collection and conservation of endangered medicinal and aromatic plants. *In: Course Compendium-Winter School on Advances in Medicinal Aromatic and Underutilized Plants Research* (eds. C. S. Tyagi, P. K. Verma, J. S. Hooda, O. P. Yadav, and R. K. Goyal) pp. 1-7. CCSHAU, Hisar, Haryana, India.
- Subedi, A., B. Kunwar, Y. Choi, Y. Dai, T. van Andel, R. P. Chaudhary, H. J. de Boer, and B. Gravendeel. 2013. Collection and trade of wild-harvested orchids in Nepal. *J. Ethnobiol. Ethnomed.*, **9**: 64
- Swarts, N. D. and K. W. Dixon. 2009a. Terrestrial orchid conservation in the age of extinction. Ann. Bot., 104: 543-56.
- Swarts, N. D. and K. W. Dixon. 2009b. Perspectives on orchid conservation in botanic gardens. *Trends Plant Sci.*, 14: 590-98.
- Tamer, C., B. Karaman, and O. Copur. 2006. A traditional Turkish beverage: Salep. *Food Rev. Inter.*, **22**: 43-50.
- Teoh, E. S. 2016. *Medicinal Orchids of Asia*. Springer International Publishing, Cham, Switzerland.

- Thakur, M. and V. K. Dixit. 2007. Aphrodisiac activity of Dactylorhiza hatagirea (D. Don) Soo in Male Albino rats. J. Evid. Based Complementary Altern. Med., 4: 29-31.
- Thammasiri, K. 2015. Current status of orchid production in Thailand. *Acta Hort.*, **1078**: 25-33.
- UNEP-WCMC. 2017. CITES Trade Statistics Derived from the CITES Trade Database. UNEP World Conservation Monitoring Centre, Cambridge. Available at: https:// trade.cites.org.
- Uniyal, S. K., A. Awasthi, and G. S. Rawat. 2002. Current status and distribution of commercially exploited medicinal and aromatic plants in upper Gori valley, Kumaon Himalaya,Uttaranchal. *Curr. Sci.*, **82**: 1246-50.
- USDA. 2016. Floriculture Crops 2015 Summary. United States Department of Agriculture, National Agricultural Statistics Service. Available at: http://usda.mannlib.cornell.edu/usda/ current/FlorCrop/FlorCrop-04-26-2016.pdf.
- van der Niet, T., R. J. Cozien, and S. D. Johnson. 2015. Experimental evidence for specialized bird pollination in the endangered South African orchid Satyrium rhodanthum and analysis of associated floral traits. Bot. J. Linn. Soc., 177: 141-50.
- van der Niet, T., D. M. Hansen, and S. D. Johnson. 2011. Carrion mimicry in a South African orchid: Flowers attract a narrow subset of the fly assemblage on animal carcasses. *Ann. Bot.*, **107**: 981-92.
- Vanlalruati, T. Mandal., and S. Pradhan. 2016. Habitat studies for conservation of medicinal orchids of Mizoram. J. Orchid Soc. India, 30: 15-18.
- Veldman, S., J. Otieno, T. van Andel, B. Gravendeel, and H. de Boer. 2014. Efforts urged to tackle thriving illegal orchid trade in Tanzania and Zambia for chikanda production. *TRAFFIC Bull.*, 26: 47-50.
- Veldman, S., B. Gravendeel, J. N. Otieno, Y. Lammers, E. Duijm, A. Nieman, B. Bytebier, G. Ngugi, F. Martos, T. van Andel, and H. de Boer. 2017. High-throughput sequencing of African chikanda cake highlights conservation challenges in orchids. *Biodivers. Conserv.*, 26: 2029-46.
- Willis, K. J. 2017. State of the World's Plants 2017 Report. Royal Botanic Gardens, Kew, London, UK.
- Wu, S. J., Y. S. Liu, T. W. Chen, C. C. Ng, W. S. Tzeng, and Y. T. Shyu. 2009. Differentiation of medicinal *Dendrobium* species (Orchidaceae) using molecular markers and scanning electronmicroscopy. *J. Food Drug Anal.*, **17**: 474-88.
- Yao, H., J. Y. Song, X. Y. Ma, C. Liu, Y. Li, H. X. Xu, J. P. Han, L. S. Duan, and S. L. Chen. 2009. Identification of *Dendrobium* species by a candidate DNA barcode sequence: The chloroplast psbA-trnH intergenic region. *Planta Med.*, **75**: 667-69.
- Yeung, E. C. 2017. A perspective on orchid seed and protocorm development. *Bot. Stud.*, 58: 33.

2018)